

## Phylogeography of foot-and-mouth disease virus in Tanzania

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## Introduction Position of Country

- ▶ Tanzania lies between 6° South and 35° East latitude and longitude respectively.
- ▶ Covers an area of 945,000 km<sup>2</sup>. and two thirds of the country land resource is rangelands.
- ▶ Tanzania borders with Kenya, Uganda, Mozambique , DRC, Rwanda, Burundi, Zambia, Malawi and Indian Ocean.
- ▶ The restrictions of livestock and wildlife movement between borders is limited.

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## Introduction–Climate

- ▶ The climate is seasonal, unimodal (Dec.–April) and bimodal (Oct.–Dec. and March–June)
- ▶ Rainfall varies widely across and between regions.
- ▶ Temp. variation (daily average) is from 25 to 31°C in dry months (Dec.–March) and 15 to 25 °C in wet months (April – July)

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## Introduction–The Disease

- ▶ FMDV, is classified within the Aphthovirus genus as a member of the Picornaviridae family.
- ▶ Foot and mouth disease (FMD) is the most contagious disease of mammals.
- ▶ Typical cases of FMD are characterised by a vesicular condition of the feet, buccal mucosa and, in females, the mammary glands.

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## Introduction–The Disease

- ▶ In Tanzania, FMD is the first most important viral TAD (MLDF, 2006).
- ▶ FMD is endemic in Tanzania since it was first documented in 1927.
- ▶ The first virus isolation and typing was done in 1954 (WRLF).
- ▶ Four common serotypes isolated and identified in Tanzania include Type A, O, SAT 1 and SAT 2.

## Objectives

- ▶ To use genetic data to determine the distribution of FMDV lineages in Tanzania.
- ▶ To find the possible sources FMDV causing outbreaks in Tanzania.

## Sample collection and preservations

- ▶ 361 epithelial tissues, probangs and sera samples from field all over Tanzania(2008–2012).
- ▶ Samples were collected from clinical animals.
- ▶ Suspended in transport media (PBS and Glycerol).
- ▶ Samples homogenized in PBS pH 7.2

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## Molecular screening/serotyping

- ▶ Extraction done using QIAamp® Viral RNA Mini Kit (50)
- ▶ Samples were tested using the Real time RT–PCR (Shaw *et. al* 2007)
- ▶ FMDV positive samples were typed (A,O C, SAT2) by one step RT–PCR using serotype specific primers (Knowles et al.,2005, 2009)

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## Results for Real Time PCR

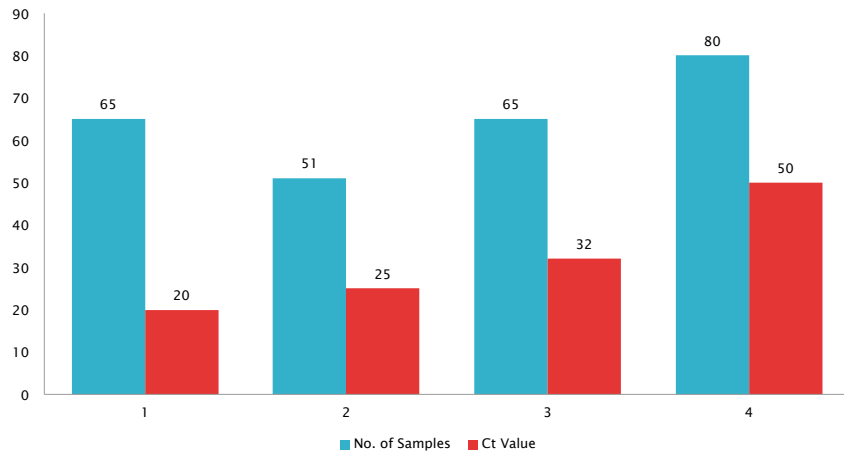
### Real Time PCR Results interpretation

- ▶ Samples with the Ct value  $<32$  were considered as **POSITIVE**.
  
- ▶ Samples with **NO** Ct value were considered as **NEGATIVE**.
  
- ▶ Samples with the Ct value  $\leq 32 - <50$  were considered as **Inconclusive**.

## Results Real Time PCR

- ▶ Samples screened were 361
- ▶ Positive samples were 181  $< 32$  as below,
  - Samples with Ct value  $<20$  were 65
  - Samples with Ct value 21–25 were 51
  - Samples with Ct value 25–32 were 65
- ▶ Samples with Ct value  $\leq 32 - <50$  were 80
- ▶ Negative samples were 100 with no Ct

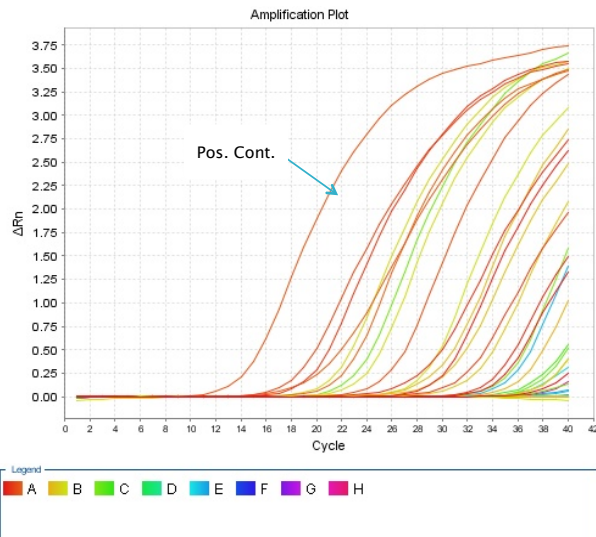
## Results Real Time PCR



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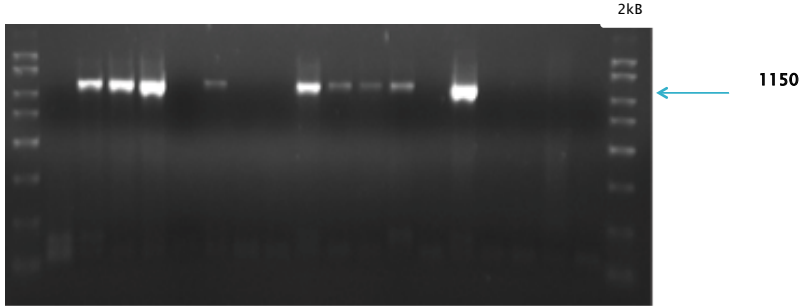
## Results Real Time PCR



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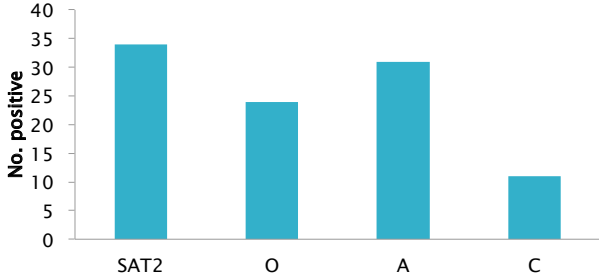
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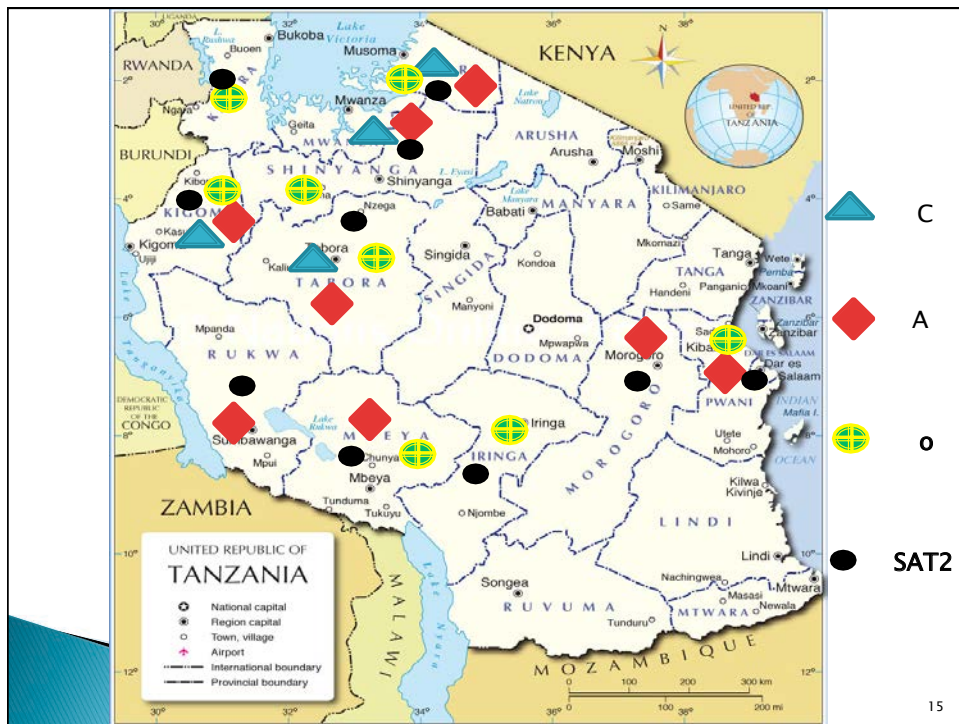
# Results Serotyping for O



# Results Serotyping

98 Samples were typed using serotype specific primers for O (24), A (32), SAT2 (34) and C?? (11)





## Discussion

- ▶ The results indicates that the 3 predominant serotypes are still exist (A, O and SAT2).
- ▶ Serotype SAT2 scored highest followed by A, O and C???? (11 samples).
- ▶ Most of outbreaks occurred in the Southern High land, Western part of Tanzania, Eastern and North west.
- ▶ No any outbreak reported in greater part of the Central Tanzania to date.



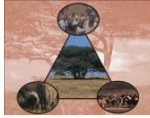
## Discussion

- ▶ Observation of multiple infections in one samples
- ▶ Climate has an effect on the recent outbreaks.
- ▶ Movement of animals to better grazing areas
- ▶ Wildlife/Livestock interface

## Future Work

- ▶ Typing serotypes SAT1 and SAT3
- ▶ Sequencing and phylogenetic analysis of the VP1 Sequences
- ▶ To check the sensitivity and Specificity of Antigen Vs. Serotyping methods.
- ▶ To complete the phylogeography study.
- ▶ Start the assay for qRT-PCR for serotype specific.

## ACKNOWLEDGEMENT



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